

1. Introduction

α2δ auxiliary subunits

α₂δ1

- Represent important therapeutic targets for gabapentinoids¹.
- Modulate high-voltage-activated voltage-gated calcium channels (VGCCs)
- \Rightarrow Enhance channel trafficking and function^{1,2}.

CACHD1

 Within the α2δ family, CACHD1 recently emerged as a modulator of low-voltage-activated (LVA) VGCCs³.

The MIDAS motif and glycosylation of CACHD1 are important for expression and function as a Ca_v3.1 voltage-gated calcium channel modulator

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MIDAS motif

How does CACHD1 modulate LVA VGCCs?

Glycosylation

- Metal ion-dependent adhesion site motif.
- Essential for the trafficking and synaptic function of α2δ and VGCCs^{1,2}.
- α2δ subunits contain fully conserved MIDAS motif (DxSxS).
- Plays crucial role in $\alpha 2\delta$ function, stability, and surface expression.
- N136 and N184 necessary for $\alpha 2\delta$ function
- Affect interaction with other subunits and VGCCs.







Fig. 1: (A) 'AAA' MIDAS motif mutation impaired CACHD1 expression at cell surface. Myc-tagged CACHD1-wt and CACHD1-G236S detected in intracellular compartments (yellow arrows) and at cell surface (white arrows), while CACHD1-AAA detected in intracellular compartments only; (scale bar 10 μ m; n=5). **(B)** Immunoreactive Myc-tagged CACHD1 proteins detected at 150-170 kDa. **(C)** Significant reduction in expression levels of CACHD1-AAA (21.3±6.8%), but not CACHD1-G236S (93.3±4.2%); (mean±SEM; One-way ANOVA, Tukey's *post hoc* test, *p<0.05, ns – non-significant; n=5).







Fig. 4: (A) Mutagenesis of predicted N-linked glycosylation sites (N145/329/373/587/940/985/Δ6) showed no effect on CACHD1 trafficking to cell surface. **(B)** Decrease in molecular mass was observed for all mutants, except N985Q. NΔ6Q mutant presented at larger molecular mass compared to CACHD1 de-glycosylated with PNGase F, suggesting additional N-linked glycosylation sites.

5. Proposed mechanism of CACHD1-Ca_v3.1 interaction



Fig. 5: Proposed mechanism of CACHD1-Ca_v3.1 interaction.

(A) CACHD1 and $Ca_V 3.1$ interact intracellularly, forming a complex that is trafficked to the membrane. CACHD1 promotes $Ca_V 3.1$ cell surface localisation³ and modulates $Ca_V 3.1$ electrophysiological properties.

(B) Mutagenesis of MIDAS motif (DxGxS to AxAxA), disrupts CACHD1 membrane trafficking, leading to intracellular retention of CACHD1/Ca_V3.1 complex,



Fig. 2: 'AAA' mutation in CACHD1 MIDAS motif prevents CACHD1-mediated rCa_V3.1 current increase and further decreases rCa_V3.1 T-type currents by 0.55-fold. **(A, B, C)** I-V relationship (holding potential -90 mV; two-way ANOVA, Bonferroni *post hoc* test, curve fitted with modified Boltzmann). **(D, E, F)** Representative current density traces at -20 mV. **(G)** Maximal conductance (two-tailed unpaired Student's *t* test); (**p*<0.05, ***p*<0.01, ****p*<0.001, *****p*<0.0001).

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References:
1 Cassidy, J. S., et al. (2014). PNAS USA, 111(24), 8979–8984.
2 Hoppa, M. B., et al. (2012). Nature, 486(7401), 122–125.
3 Cottrell, G. S., et al. (2018). Journal of Neuroscience, 38(43), 9186–9201.

resulting in reduction of Ca_v3.1 T-type currents.

6. Conclusions

 The MIDAS motif plays an important role in CACHD1-mediated increase of T-type currents in Ca_v3.1 VGCCs.

 Mutagenesis of the MIDAS motif significantly reduces CACHD1 expression and cell surface trafficking, similar to effects seen in α2δ subunits.

 Glycosylation does not appear to have a key role in CACHD1 trafficking, and further investigation is needed to assess its effect on CACHD1 function

 CACHD1 represents a promising therapeutic target for modulating Ca_V3 VGCCs in hyperexcitability conditions (e.g., pain, epilepsy), similar to gabapentinoid targeting of α2δ subunits.