

Development of an Impedance Based Screening Assay

for Cardiac Safety and Cardiotoxicity Detection in Stem Cell-derived Cardiomyocytes



Robert W. Kirby, Said El Haou, Edward Humphries, John Ridley and Marc Rogers

Metrion Biosciences Ltd, Riverside 3, Granta Park, Cambridge CB21 6AD, U.K.

Introduction

Cardiac toxicity remains the leading cause of new drug safety side-effects. Current preclinical cardiac safety assays rely on *in vitro* cell-based ion channel assays and *ex vivo* and *in vivo* animal models⁽¹⁾. These assays provide an indication of acute risk but they do not always predict the effect of chronic compound exposure, as recently seen with oncology drugs⁽²⁾. Therefore, new assays are required to characterise chronic structural and functional effects in human cells earlier in drug discovery^(2,3). Impedance-based technology can provide more accurate chronic cardiotoxicity measurements in an efficient manner using human induced pluripotent stem cell-derived cardiomyocytes (hiPSC-CMs)^(1,3).

Here we outline validation of chronic cardiotoxicity assays using an impedance platform in combination with commercial hiPSC-CMs:

1. Impedance readouts can be reliably measured and are stable to allow acute (< 24 hrs) as well as longer duration (72 hrs) testing.
2. Identify the acute and chronic effects of clinical cardiac ion channel modulators against cardiomyocyte contractility and viability.
3. Confirm that structural and functional cardiotoxicity of clinical compounds arising from multiple cellular and signalling mechanisms could be detected in an acute and/or chronic assay timeframe.

Materials and methods

hiPSC-CMs from commercial vendors (Ncardia and CDI) were plated onto CardioExcyte96 well plates (Nanion) according to manufacturers' instructions. Plates were incubated at 37 °C (5 % CO₂) for 7-10 days and 100 % medium exchanges were performed daily. Impedance signals were recorded using the CardioExcyte96 (CE96) platform and the formation of a spontaneous beating syncytial network was monitored with 30 s impedance recording every hour (1 hr) during the first week in culture.

A single concentration of compound was then applied to each well (N ≥ 4) every 24 hrs as a 1:1000 fold dilution into media (final = 0.1 %) for a maximum of 72 hours before a washout period. The outputs (Figure 1) were recorded at 60 minute intervals to assess acute vs. chronic effects (< 24 hrs vs 24 – 72 hrs).

All data are presented as mean ± S.D. Parameter values were normalised to the baseline (pre-drug) and presented as percent of control.

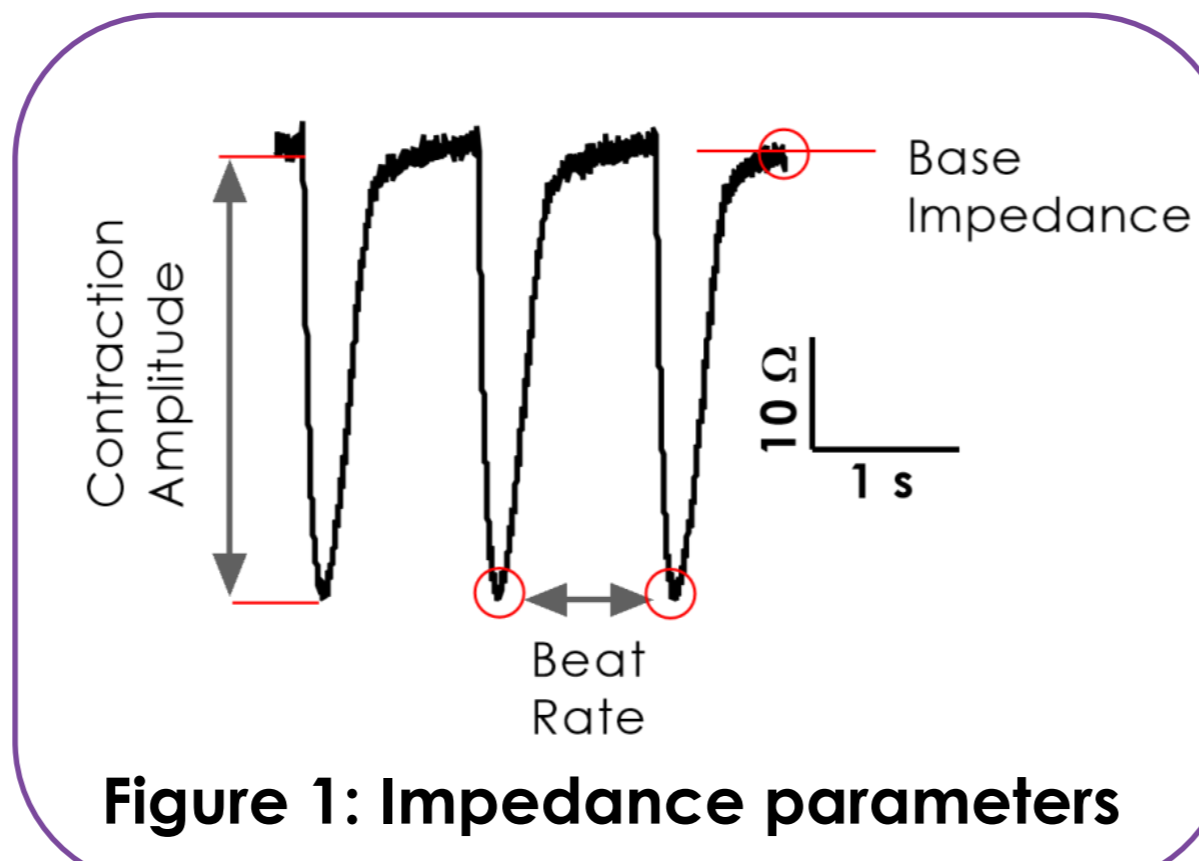
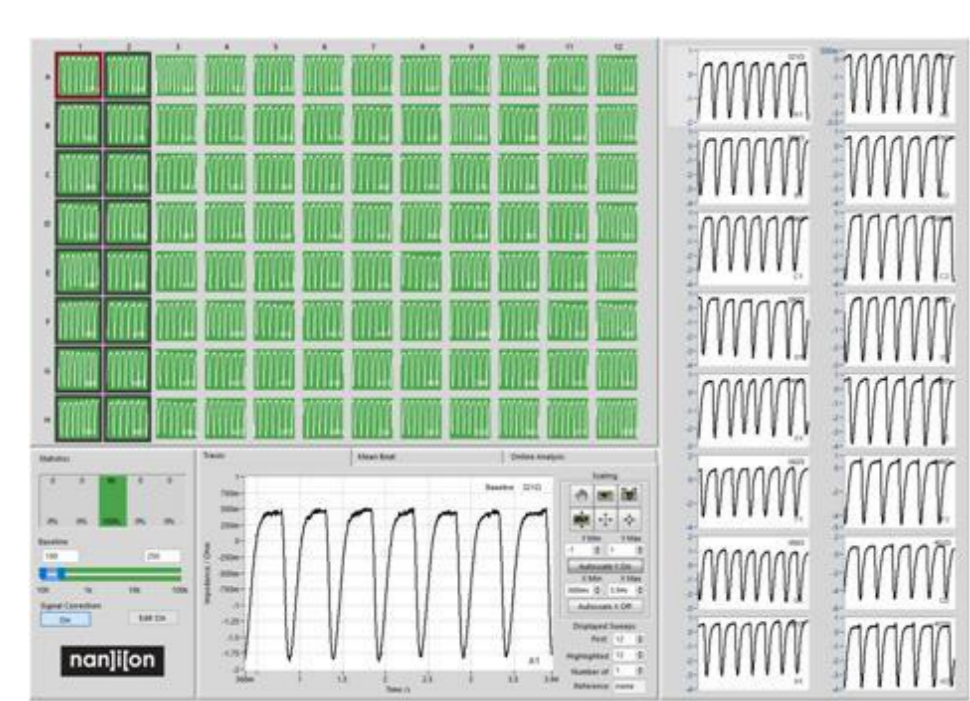


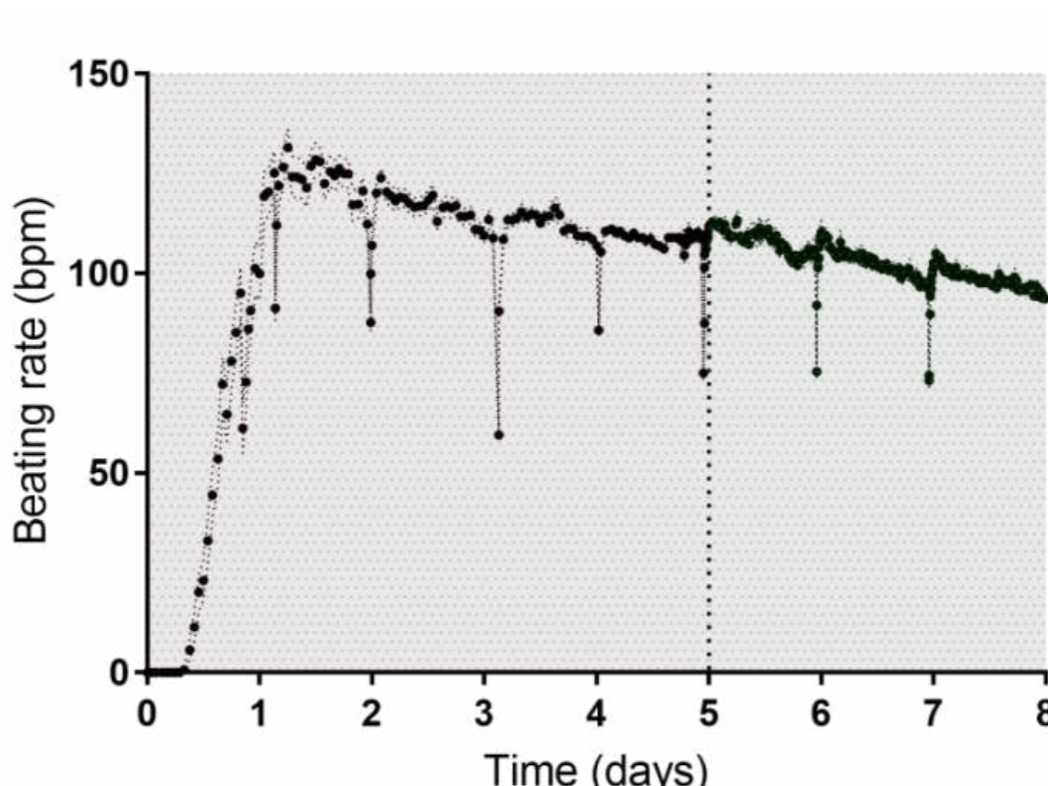
Figure 1: Impedance parameters

1. Impedance assay

A. CE96 data window



B. Beat rate



C. Base impedance

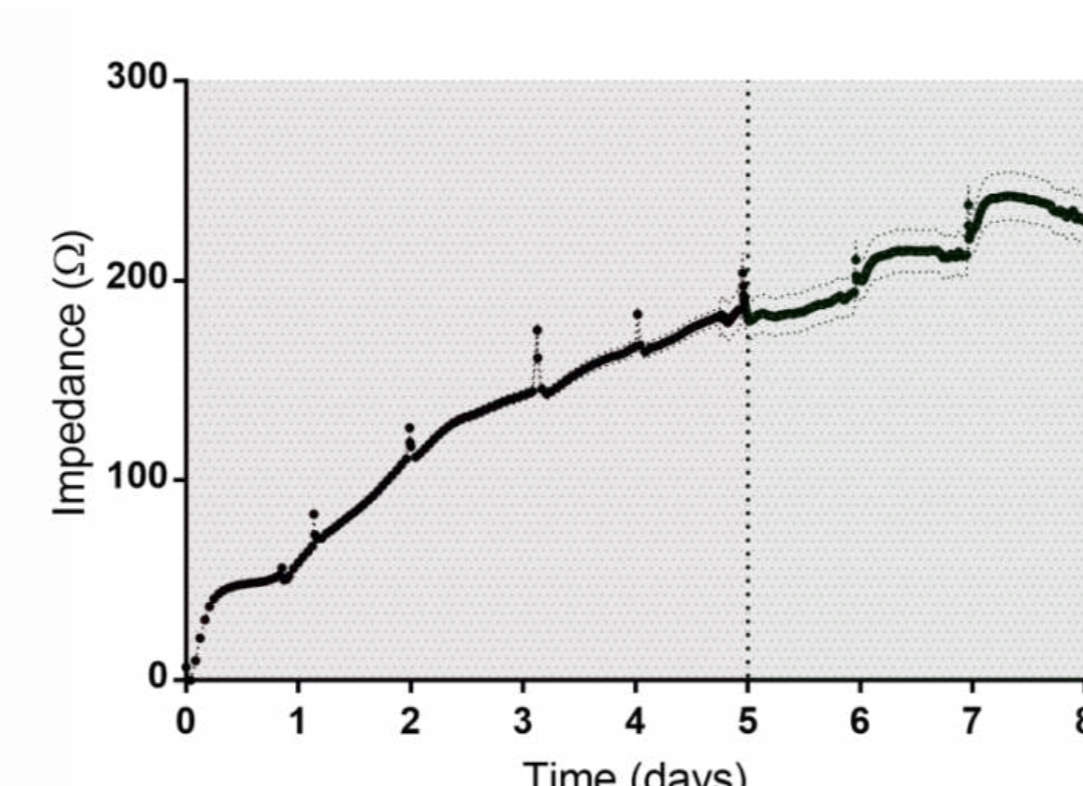
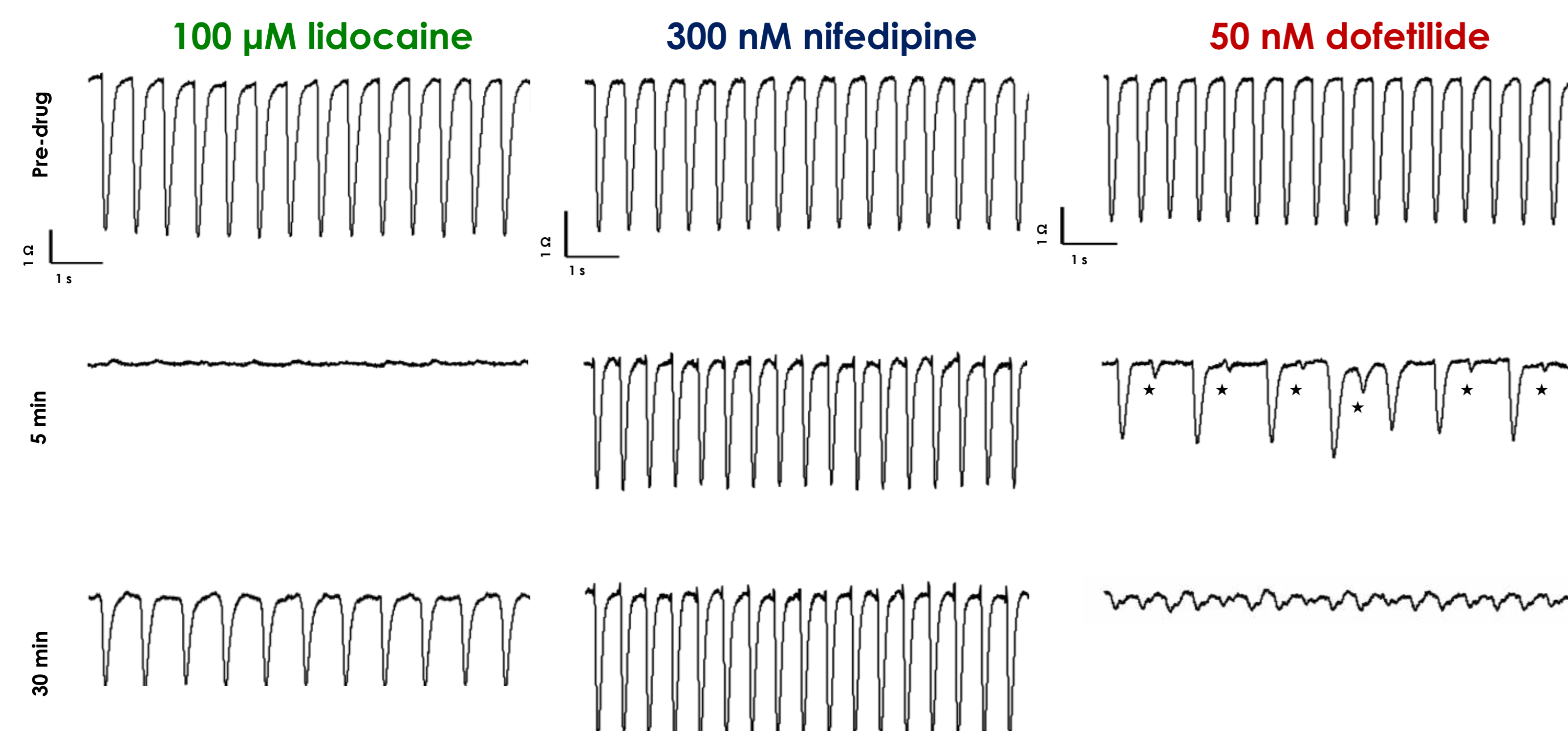


Figure 2: Representative baseline impedance data recorded from CE96

hiPSC-CMs from a commercial vendor plated onto a CardioExcyte96 well plate delivered a high success rate across the plate (A) with >90% of wells beating within 6 hours that stabilised within 72 hours (B). Consistent base impedance (growth phase) parameters were obtained after 5 days (C) that were suitable for chronic drug screening.

2. Cardiac ion channel effects on contractility

A. Acute effects on cell contraction



B. Chronic cardiotoxicity effects

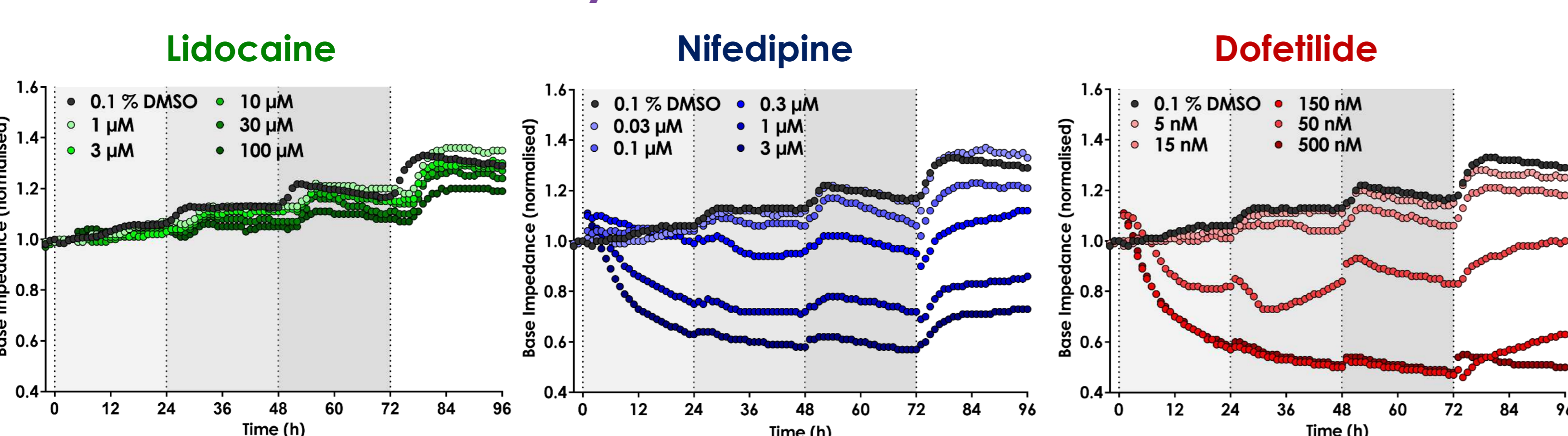


Figure 3: Acute cardiac effects of I_{Kr}, I_{Na} and I_{Ca} ion channel modulators on cell contraction

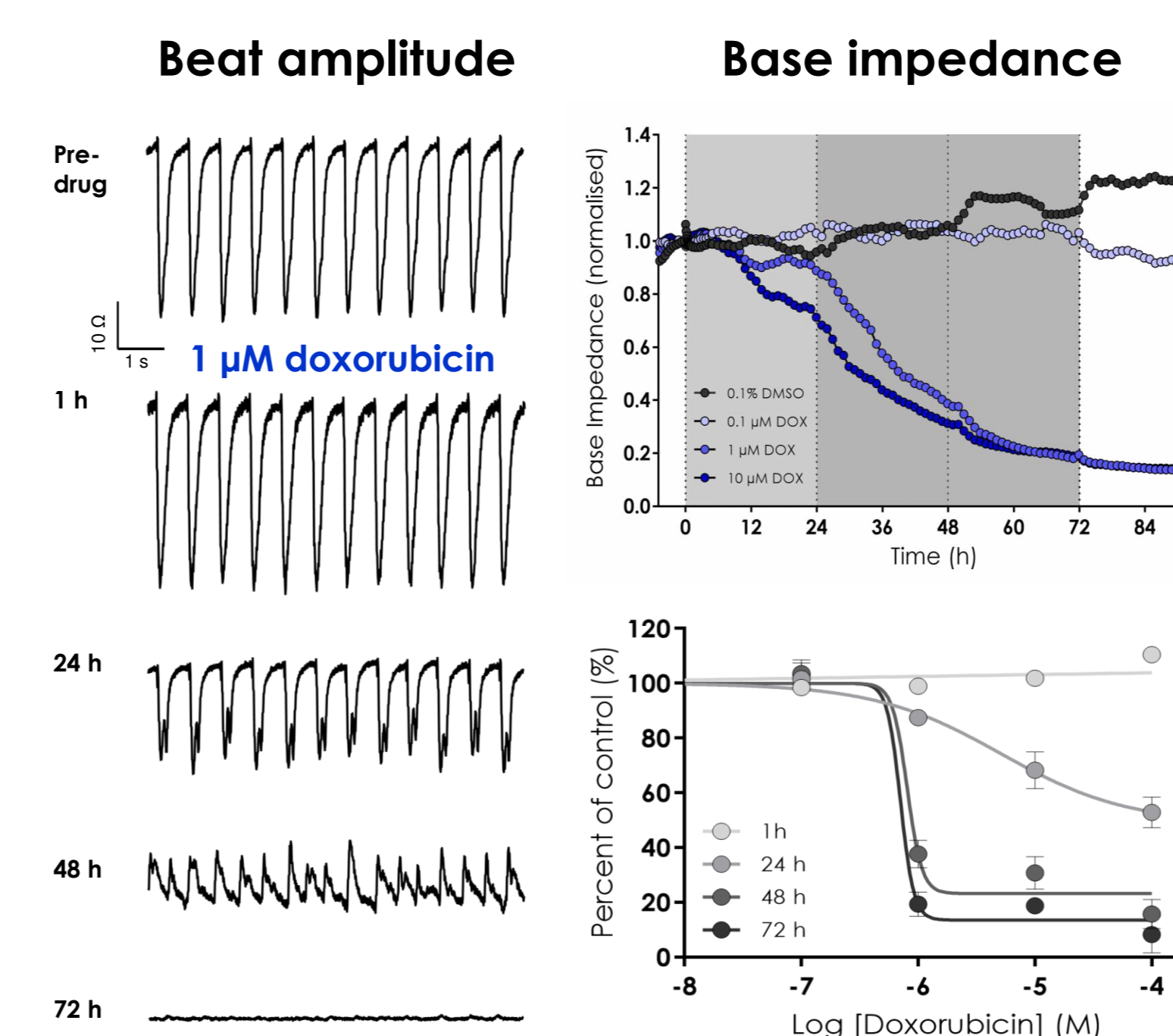
(A): Cardiac contractility was assessed by examining changes in beat rate and amplitude and the expected acute effects of cardiac ion channel modulators of I_{Kr} (dofetilide, 50 nM), I_{Na} (lidocaine, 100 μM) and I_{Ca} (nifedipine, 300 nM) were confirmed as represented by the traces above showing spontaneous contractile activity in control conditions (Pre-drug) and in presence of drug after 5 and 30 min incubation. Lidocaine stopped contraction but recovered with a decreased beat rate & amplitude. Nifedipine increased beat rate but decreased amplitude and dofetilide decreased amplitude & beat rate with secondary beats degenerated into fibrillation. ★: secondary beat

(B): **Chronic cardiac drug effects on base impedance**
Base impedance was used to monitor cell viability over 72 hour period. Lidocaine had no effect at 72 hrs relative to control whereas nifedipine and dofetilide produced time- and concentration-dependent decreases in base impedance within 24 hrs.

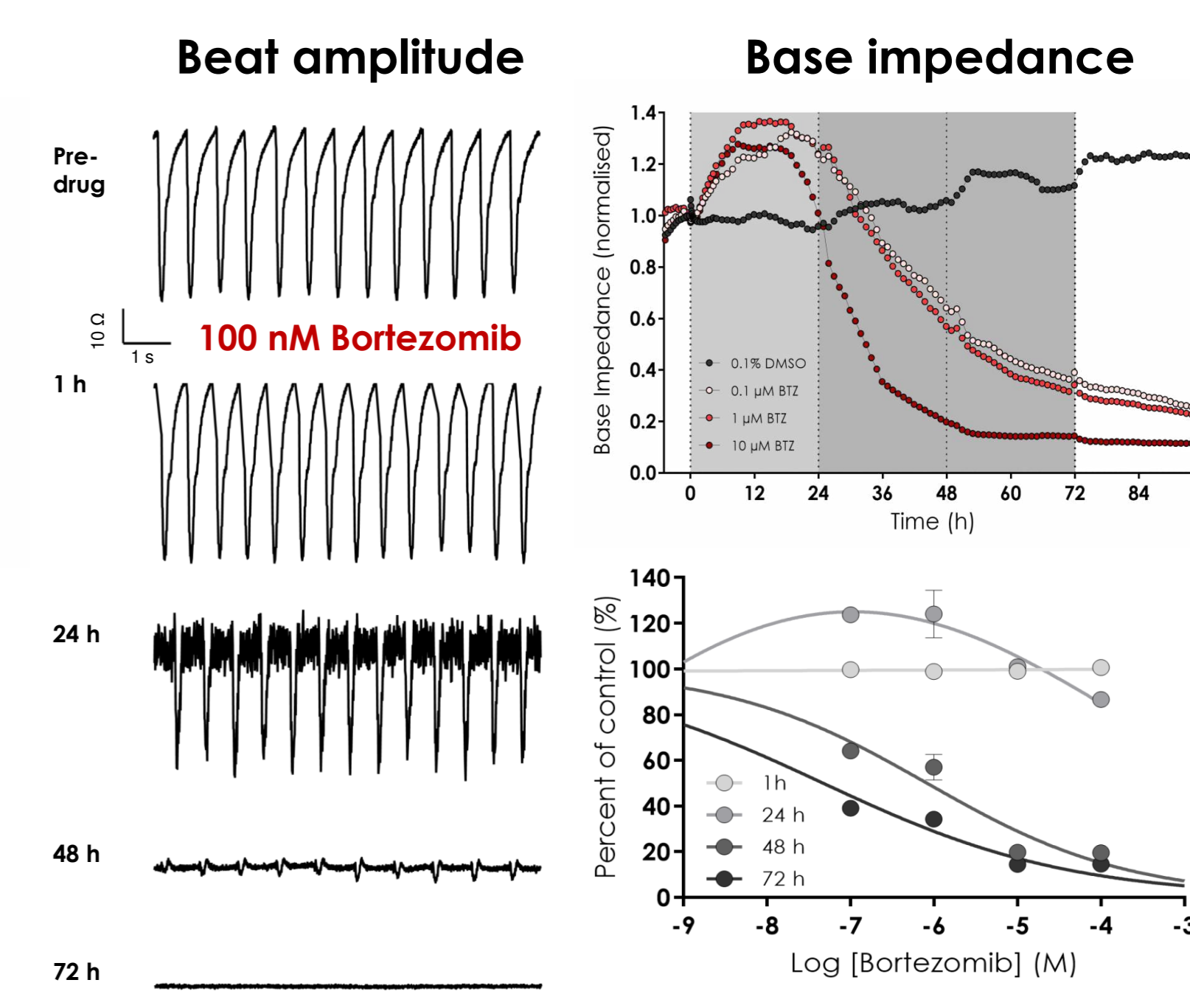
3. Chronic cardiotoxicity mechanisms

Functional and structural cardiotoxic effect of drugs with different modes of action, including several therapeutic oncology reagents, were tested. Drug mechanisms included DNA intercalation (doxorubicin), proteasome inhibition (bortezomib), myosin disruption (blebbistatin), SERCA Ca²⁺ pump inhibition (thapsigargin), kinase inhibition (Sunitinib), and an ion channel modulator with mixed functional and structural effects (amiodarone). Aspirin is considered non-cardiotoxic and was used as a negative control⁽⁴⁾.

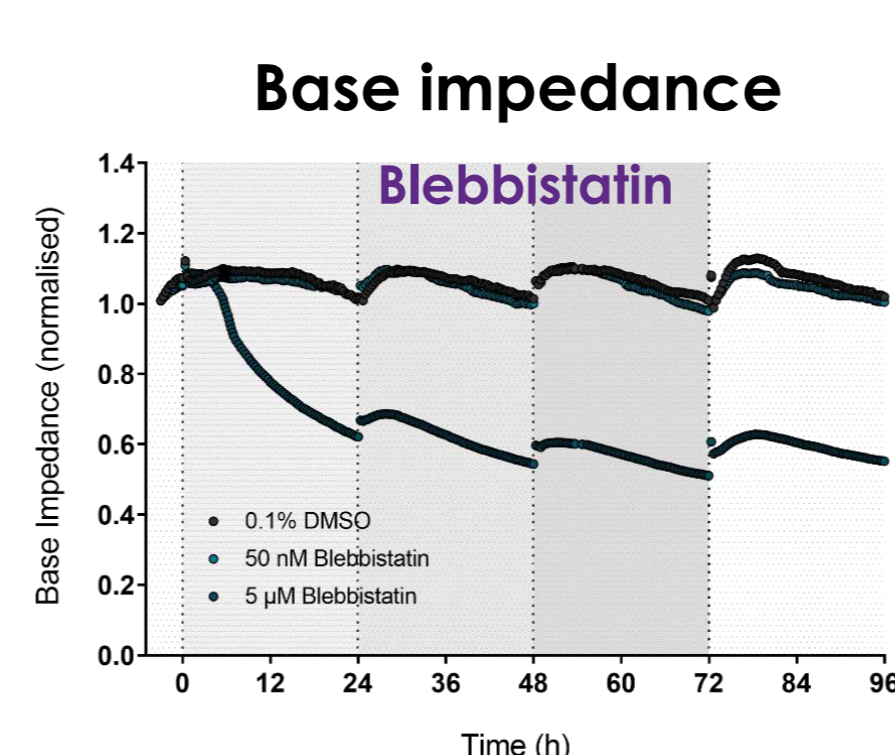
A. DNA intercalation



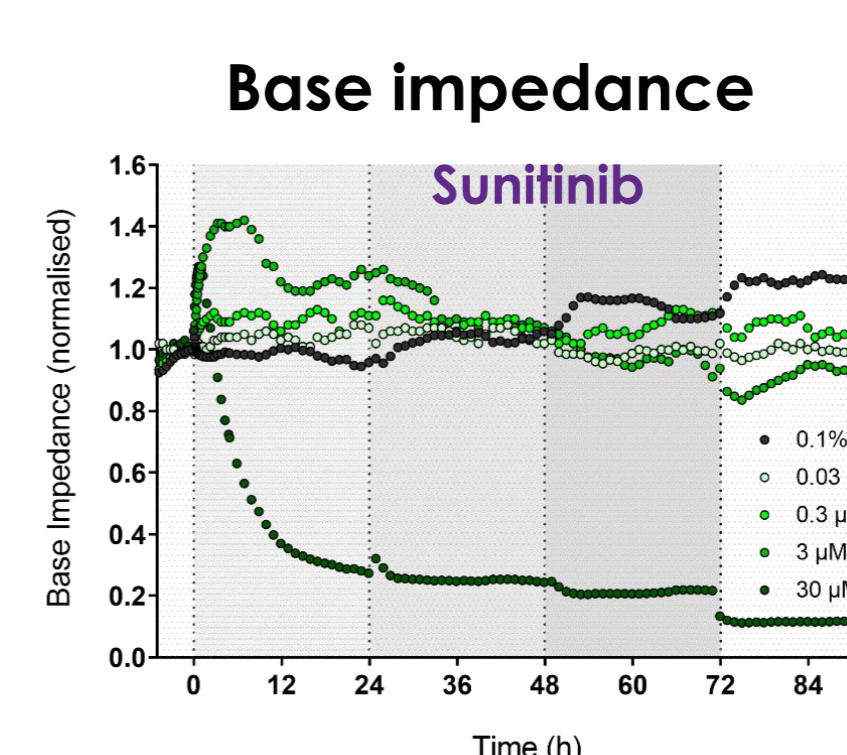
B. Proteasome inhibition



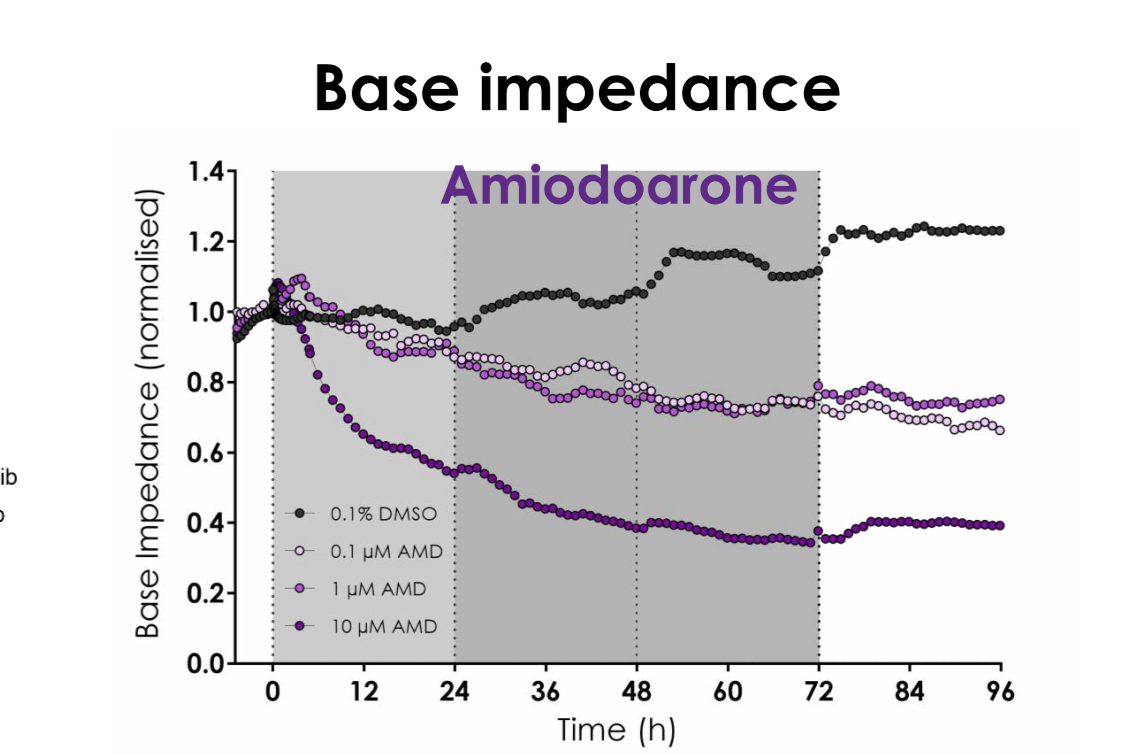
C. Myosin disruption



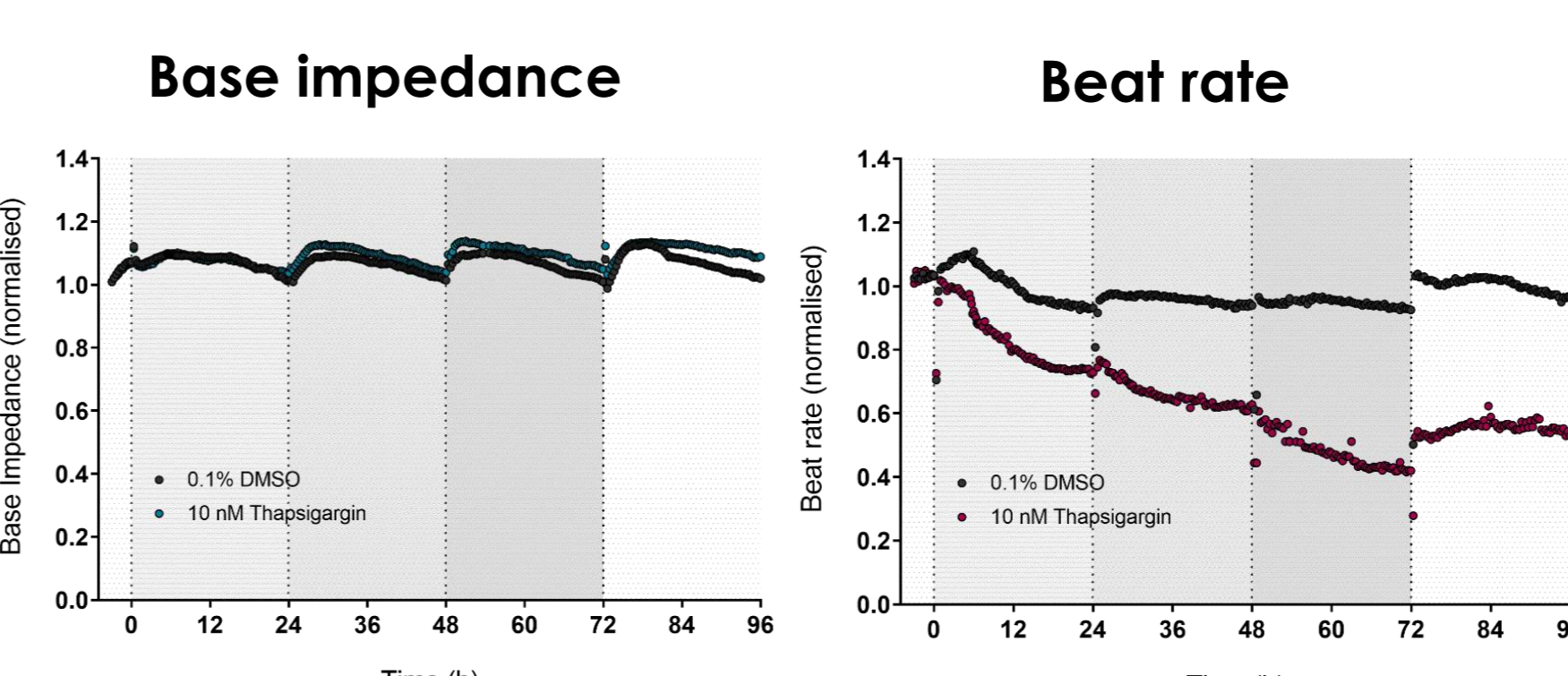
D. Kinase inhibition



E. Structural and functional



F. SERCA Ca²⁺ pump inhibition



G. Aspirin

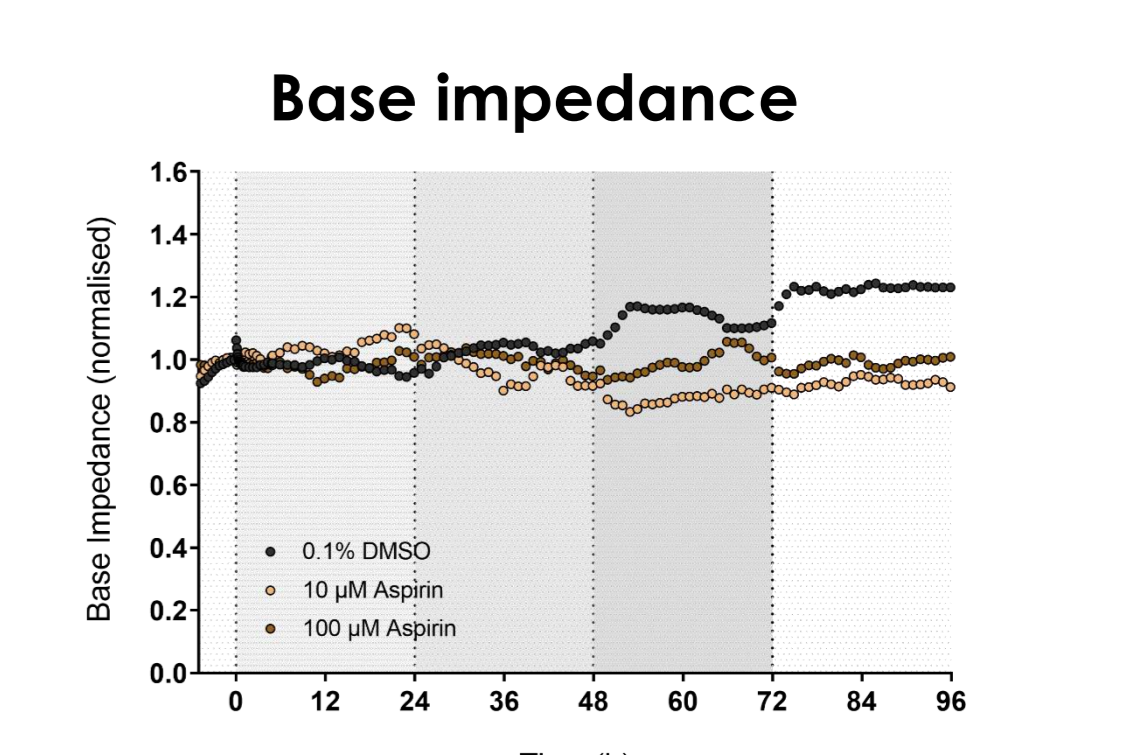


Figure 4: Examination of different modalities leading to chronic cardiotoxicity

Concentration and time-dependent decreases in base impedance were observed for doxorubicin with > 24 hrs required to resolve the full effect (A). Bortezomib produced biphasic effects, initially increasing base impedance (< 24 hrs) at all concentrations tested, before impairment of contractility was seen at > 24 hrs (B). The myosin disruptor blebbistatin produced a rapid and sustained reduction in base impedance within 24 hrs (C). Similar to bortezomib, the oncology kinase inhibitor sunitinib produced a transient and dose-dependent increase in base impedance at 3 and 30 μM before longer incubation and higher concentrations disrupted base impedance (D). The non-specific ion channel modulator amiodarone also produced concentration and time-dependent deficits in cell impedance (E). The Ca²⁺ pump inhibitor thapsigargin showed a small tendency to increase base impedance, and as expected more pronounced effects on beat rate contractility due to depletion of intracellular calcium stores (F). The negative control Aspirin showed little concentration or time-dependent effects on cell impedance (G).

H. Atypical cardiac toxicity of Trastuzumab

Trastuzumab is a monoclonal antibody approved for treating HER2-positive breast and gastric cancer. However, clinical incidents and post-market analysis reveal that therapeutic use has been limited by the occurrence of adverse cardiac events, the mechanism(s) of which are not fully realised⁽⁵⁾.

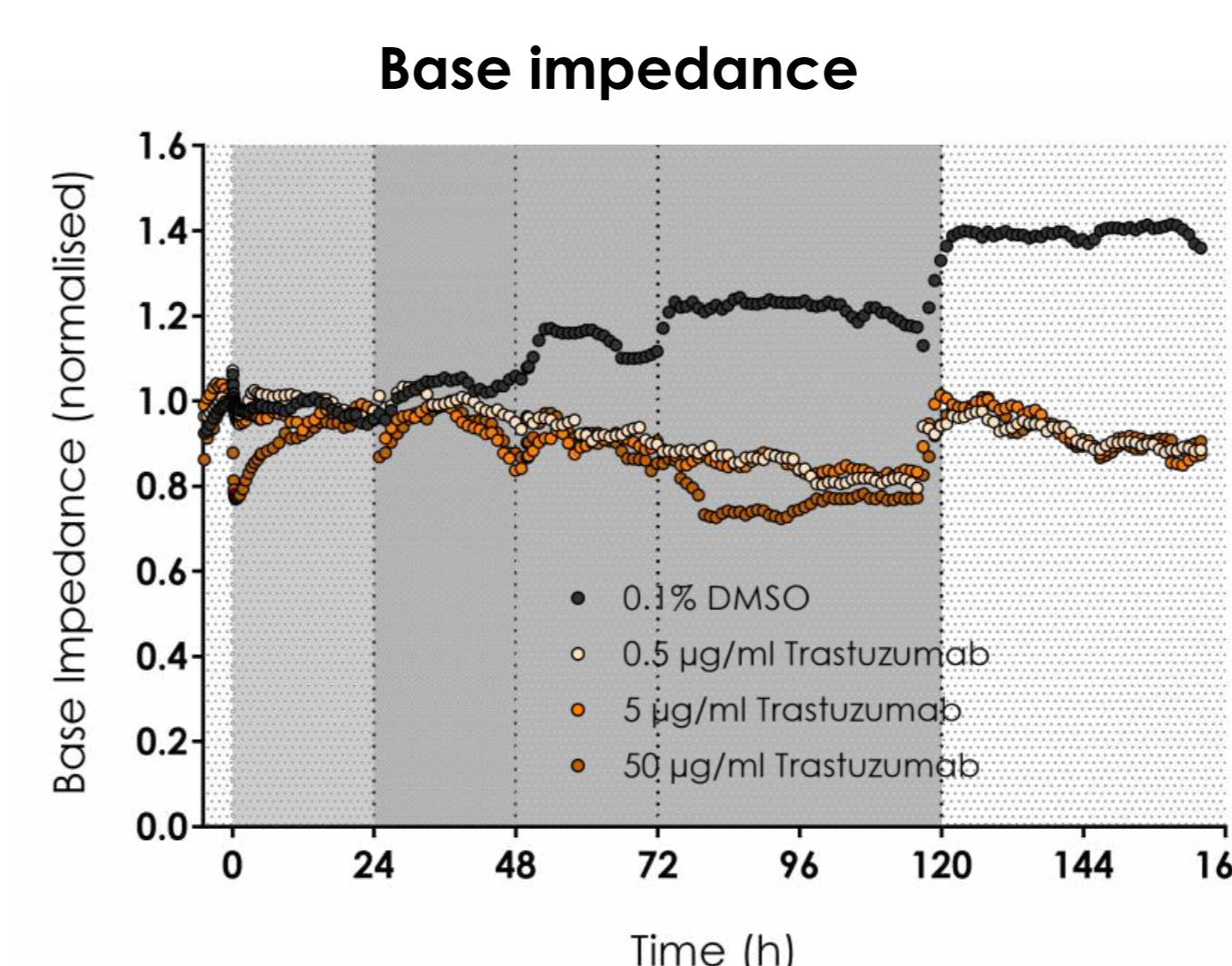


Figure 5: Trastuzumab (0.5 – 50 μg/ml) was applied at 0, 24, 48 and 72 hrs after base impedance stability was achieved. Significant effects on base impedance relative to control were only recorded > 72 hrs following multiple applications of antibody, suggesting either that entry into the cell is limited or the modulation of substrates responsible for this cardiotoxicity is slow-acting.

Conclusions

- The combination of CE96 impedance platform and commercial human stem cell-derived cardiomyocytes delivers a stable and consistent assay with a high success rate across every 96 well plate, enabling the reliable screening of acute and chronic cardiotoxicity effects of test compounds.
- Surprisingly, we detected chronic functional and structural effects of known cardiac ion channel modulators, in addition to their expected acute effects on cell excitability and contraction.
- This assay platform can reliably detect the chronic cardiotoxicity effects of clinical compounds with diverse mechanisms-of-action, indicating its utility as part of a translational cardiac safety testing regime.

Acknowledgements

Authors would like to thank former Metrion cell culture manager Louise Webdale and Metrion summer intern student Luka Smalinskaite for their assistance with the experiments.

References

- (1) Ferdinandy *et al.*, (2019). European Heart Journal. PMID: 29982507
- (2) Scott *et al.*, (2014). Toxicological Sciences. PMID: 23237062
- (3) Koci *et al.*, (2017). Toxicology and Applied Pharmacology. PMID: 28546047
- (4) Clements *et al.*, (2015). Toxicological Sciences. PMID: 26259608
- (5) Mohan *et al.*, (2018). Antibody Therapeutics. PMID: 30215054